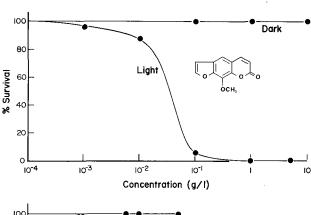
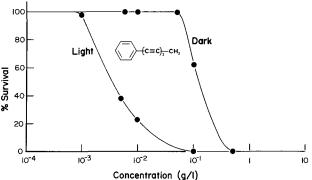
Further work will ascertain whether all 3 compounds operate through the same mechanism, and whether DNA is the target of the photosensitized reactions described in this

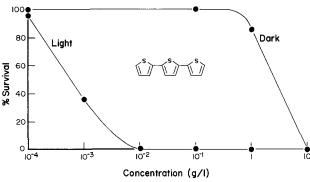
The ovicidal properties of natural products, both in dark reactions and in the presence of light, may be an important part of the plant-insect relationship. The results described here with *D. melanogaster* suggest the need for experiments in which egg survival should be scrutinized, particularly during the interaction between herbivorous insects and plants and their constituents in a natural environment.

- The help provided by Prof. Eliot B. Spiess, Department of Biological Sciences, and the financial assistance of the National Institutes of Health (GM 24144) are gratefully acknowledged.
- Kawazu, K., Ariwa, M., and Kii, Y., Agric. Biol. Chem. 41
- Nakajima, S., and Kawazu, K., Agric. Biol. Chem. 41 (1977)
- Nakajima, S., and Kawazu, K., Agric. Biol. Chem. 44 (1980) 1529.
- Goldman, A.S., and Setlow, R.B., Exp. Cell Res. 11 (1956) 5 146.
- PHT: Arnason, T., Wat, C.K., Downum K., Yamamoto, E., Graham, E., and Towers, G.H.N., Can. J. Microbiol. 26 (1980) 698; aT: Arnason, T., Chan, G.F.Q., Wat, C.K., Downum, K., and Towers, G.H.N., Photochem. Photobiol. 33 (1981) 821; 8-MOP: Song, S. P., and Tapley, Jr, K. J., Photochem. Photobiol. 29 (1979) 1177, and references cited therein.
- Parrish, J.A., The Science of Photomedicine. Eds J.D. Regan and J.A. Parrish. Plenum Press, New York 1981.
- Berenbaum, M., Science 201 (1978) 532
- Berenbaum, M., Ecology 62 (1981) 1254. Granger, M., Toulme, F., and Helene, C., Photochem. Photobiol. 36 (1982) 175, and references cited therein.

0014-4754/83/040402-02\$1.50 + 0.20/0© Birkhäuser Verlag Basel, 1983







The ovicidal activity of PHT, α T, and 8-MOP. The survival of D. melanogaster is expressed as a function of the concentration of sensitizer used, plotted on a logarithmic scale. The concentrations in µg/cm² can be obtained by multiplying the values shown by

Antifeedant activity of some ajugarin derivatives in three lepidopterous species

R. B. M. Geuskens, J. M. Luteijn and L. M. Schoonhoven¹

Department of Animal Physiology and Department of Organic Chemistry, Agricultural University, Haarweg 10, NL-6709 PJ Wageningen (The Netherlands), July 26, 1982

Summary. The trans decalin unit of clerodendrin has antifeedant activity in larvae of Pieris brassicae, but not in larvae of Spodoptera littoralis and S. exigua. It is suggested that antifeedant activity of clerodanes for Spodoptera spp. is based on the combination of a furofuran ring and epoxy diacetate groups in the decalin unit.

Naturally occurring antifeedants may occasionally be rendered much more deterrent to insects by minor modifications of their molecular structure. Thus, the antifeedant

activity of clerodendrin (fig., A), a bitter principle isolated from the Indian bhat tree (Clerodendron infortunatum), was raised 15-fold by transforming it into its methanol adduct² when Spodoptera litura was used as experimental insect. On the basis of this observation Kojima et al.3 suggested that the perhydrofuro-2,3-b-furan ring represents the active part of the clerodendrin molecule. However, none of 25 perhydrofuro-2,3-b-furan derivatives could reach the antifeedant level of the complete clerodendrin molecule³. Alternatively, the antifeedant activity could reside in the decalin unit of the clerodane molecule. This supposition is corroborated by the finding that ajugarin (fig., B), containing the same trans decalin unit, inhibits feeding in Spodoptera exempta and S. littoralis⁴. To test this hypothesis 2 stereoisomers of the decalin moiety (fig., C and D) as well as the cyclohexane derivative (E) have been synthesized⁵ and were tested for their antifeedant activity. The present report describes the effect of C, D and E on food intake in an oligophagous lepidopterous species (Pieris brassicae L.) and 2 polyphagous insects (Spodoptera littoralis and S. exigua).

Newly moulted 5th instar larvae, raised on artificial diets, were tested on foamed polystyrene (p15) lamellae 0.7 mm thick⁷. The compounds to be tested were dissolved in ethanol 96%. Each solution was mixed with an equal volume of a 0.25 M sucrose in water solution, which served as a feeding stimulant. Each lamella, measuring 18 cm², was treated with 0.25 ml of the test solution, dried for 30 min at 50 °C (C-treated lamellae were dried for 24 h at room temperature) and offered to a single insect for 24 h. The insects were starved for 3 h before the beginning of the experiments. During the tests they had access to free water. After the tests the remains of the lamellae were dried again for 30 min (at 50 °C or room temperature) and weighed. Each compound was tested in a nonchoice situation and at various concentrations. Each concentration was tested on 20 larvae. Each control series involved 10 larvae.

As shown in the table compound C inhibits feeding in P. brassicae, but not in the 2 Spodoptera species, even when tested at high concentrations. P. brassicae responds also to compound D, albeit only at a 10-fold higher concentration. Compound E, tested only in P. brassicae, is inactive, despite the presence of the spiro epoxide and one acetate group. Interestingly compound C fails to inhibit feeding in S. littoralis, whereas ajugarin shows complete feeding inhibition in this species at 300 ppm⁴. S. littoralis is known to be inhibited by clerodendrin⁶. Obviously the presence of a butanolide containing side chain is required to extend the anti-feeding activity of the epoxy diacetate group.

The following conclusions may be drawn. P. brassicae larvae react to low concentration of compound C, since 95-100% feeding inhibition is obtained at concentrations where 1 molecule of antifeedant neutralizes the feeding stimulating effect of 370 molecules of sucrose. Furthermore, it is interesting to note that the trans decalin unit, lacking the methyl group at C-8 (see F), shows anti-feeding activity in Locusta migratoria. A 70% inhibition has been observed at a concentration of F of 100 ppm⁸.

It is interesting to note that the oligophagous species P. brassicae is inhibited by 2 substances which do not affect

Antifeedant activities for compounds C, D and E

	Concentration (ppm)	1000	500	100	50	25
C	P. brassicae S. littoralis S. exigua	+ + + + 0 0	++++	++++	+++	++
D	P. brassicae S. littoralis	++++	+++			
E	P. brassicae		0			

Degree of antifeedant activity: + + + + = 100-95%; + + + = 95-75%; + + = 75-50%; + = 50-25%; 0 = 25-0%.

A Clerodendrin A; B ajugarin I; C and D stereoisomers of decalin moiety; E cyclo hexane derivative, and F decalin derivative.

OAc

F

OAc

OAc

C

the Spodoptera species tested. This would fit the idea that food specialists are more sensitive to antifeedants than generalists⁹. On the other hand, Kato et al.² have reported that clerodendrin inhibits the polyphagous larvae of S. litura from feeding, but not 3 other insect species which are monophagous. These observations stress the fact that different species may show different reactions to certain substances. This agrees with the finding that deterrent receptors in various insects show large interspecific variations 10

In conclusion it may be envisaged that the antifeedant activity of the clerodanes to Spodoptera spp. is neither located exclusively in the furofuran unit of the molecule, as Kojima and Kato³ conjectured, nor in the decalin moiety. The present results support the view that the furofuran ring and the epoxy diacetate groups of the trans decalin act synergistically, thus producing the striking antifeedant effect of clerodendrin in, for instance, S. litura.

- To whom reprint requests should be addressed.
- Kato, N., Takahashi, M., Shibayama, M., and Munakata, K., Agric. Biol. Chem. 36 (1972) 2579
- Kojima, Y., and Kato, N., Agric. Biol. Chem. 44 (1980) 855. Kubo, I., Lee, Y.W., Balogh-Nair, V., Nakanishi, K., and
- Chapya, A., J. chem. Soc. chem. Commun. 1976, 949
- Luteijn, J. M., and de Groot, A., J. org. Chem. 47 (1981) 3448.
- Wada, K., and Munakata, K., Agric. Biol. Chem. 35 (1971) 115.
- Meisner, J., Ascher, K.R.S., and Kamhi, J., Wld Rev. Pest Contr. 9 (1970) 104. Ley, S.V., Simpkins, N.S., and Whittle, A.J., J. chem. Soc.
- chem. Commun. 1981, 1001.
- Jermy, T., Ent. exp. Appl. 9 (1966) 1.
- Schoonhoven, L. M., Ent. exp. Appl. 31 (1982) 57.

0014-4754/83/040403-02\$1.50+0.20/0© Birkhäuser Verlag Basel, 1983